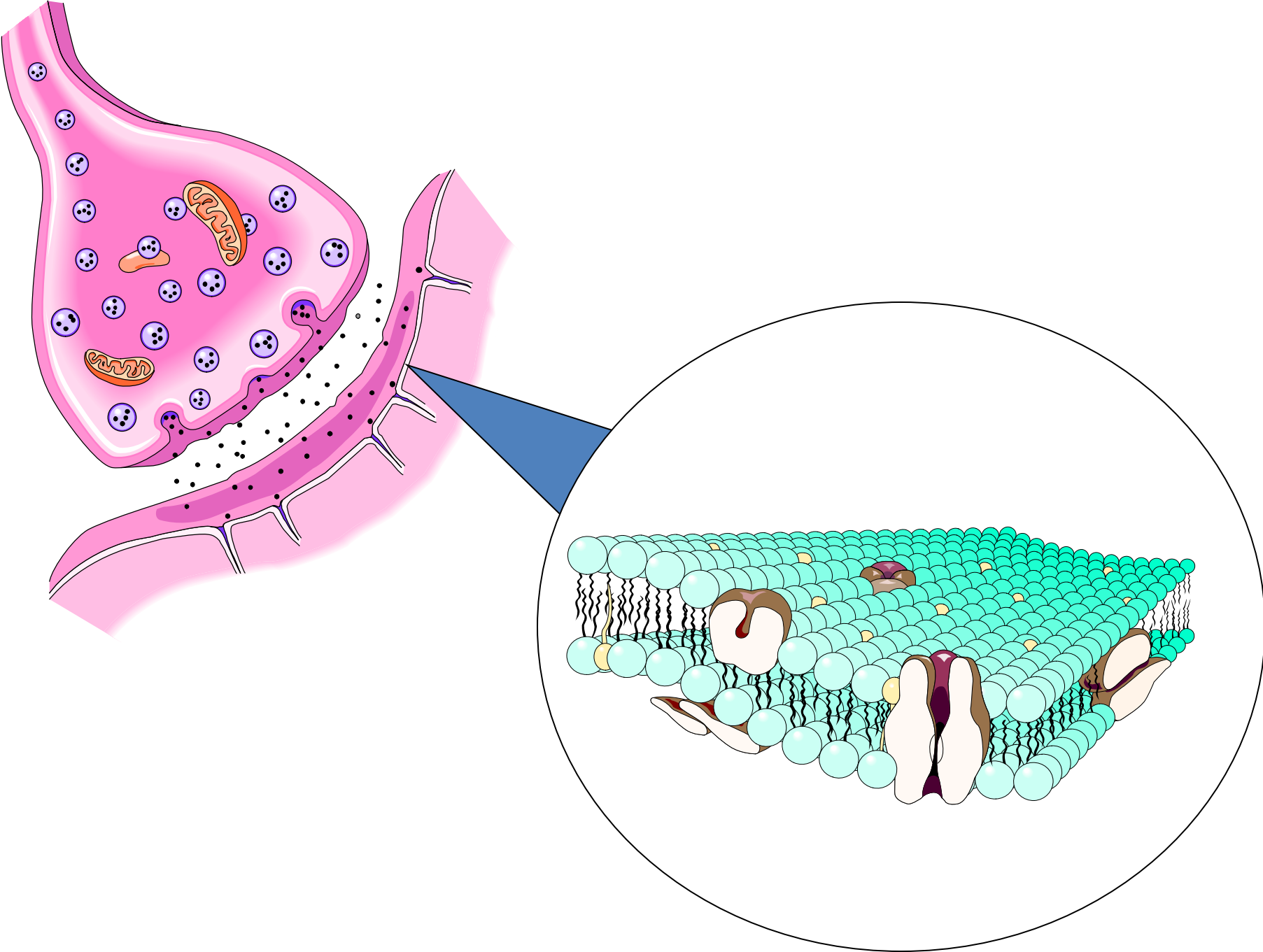


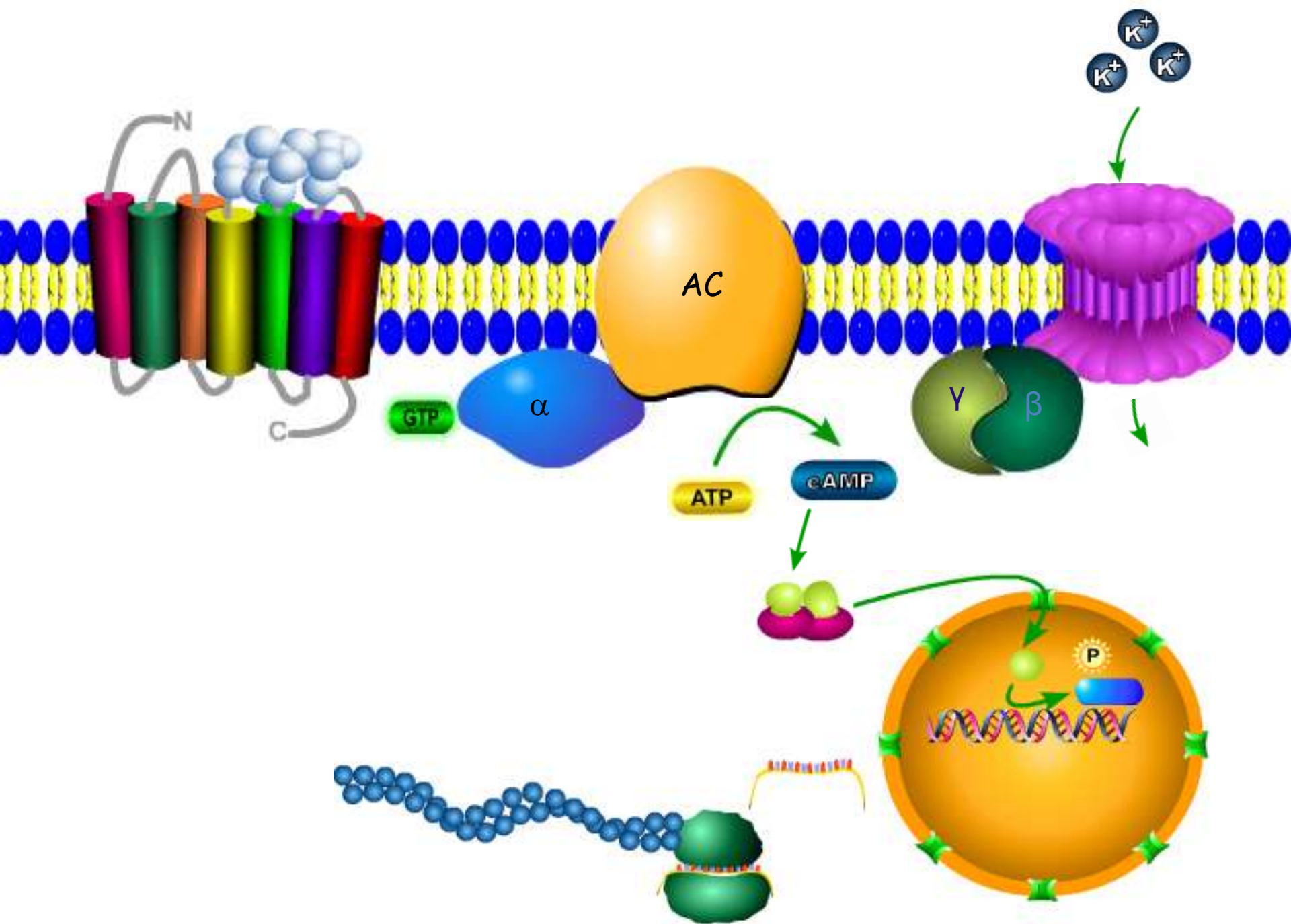
Application of baculovirus technology for Systems Biology studies of G Protein-Coupled receptors

Sergei Kopanchuk

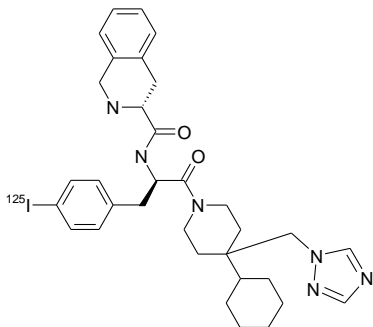
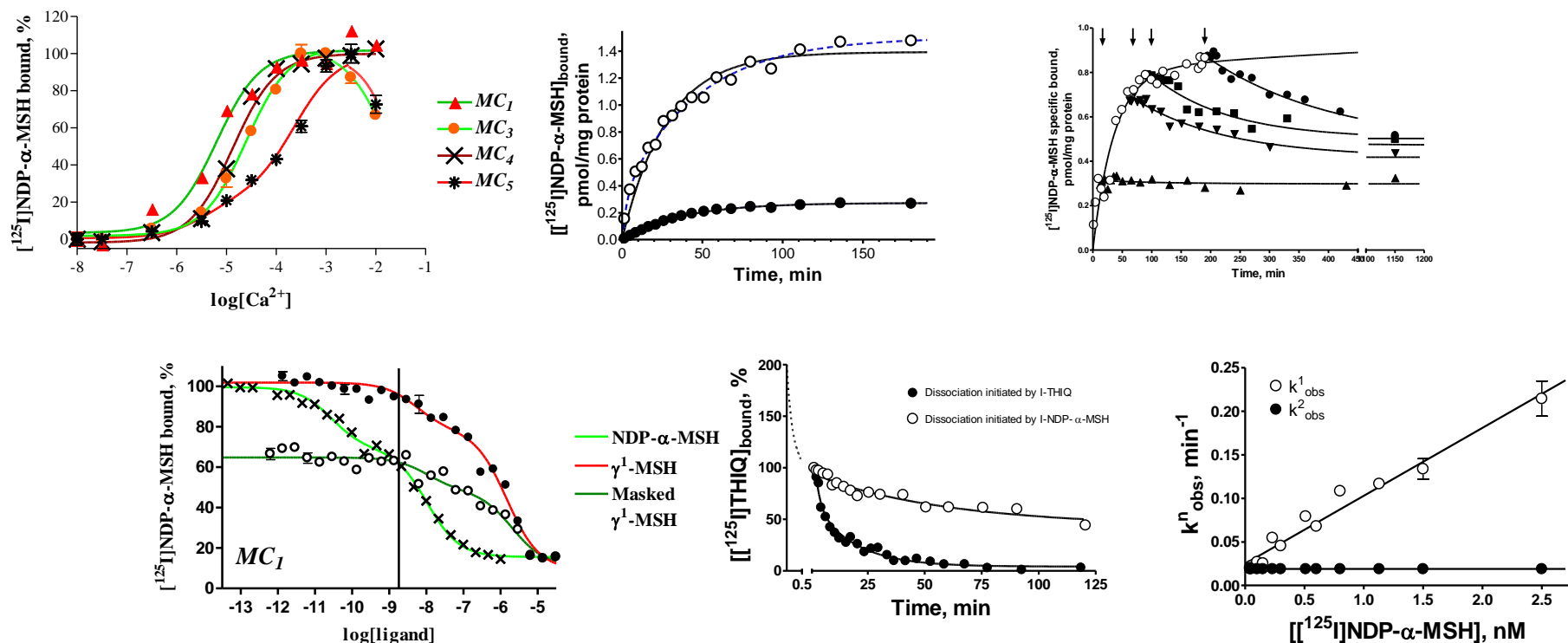
*Institute of Chemistry
chair of bioorganic chemistry*







Binding assays with radioactively labeled ligands to membranes of Sf9 cells expressing the recombinant human Melanocortin (MC) receptor subtypes

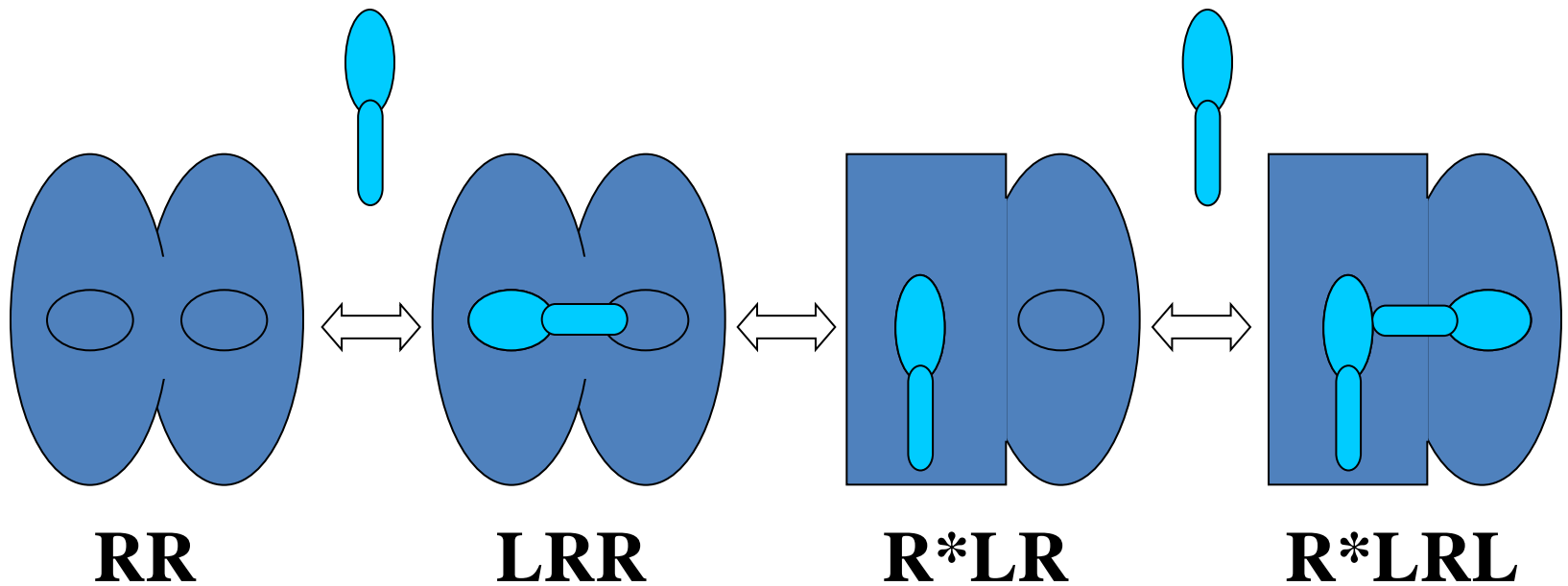
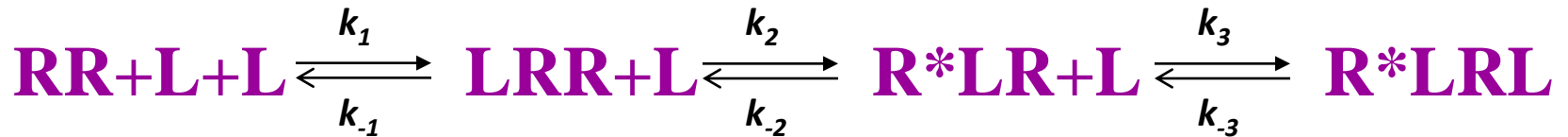


Kopanchuk S., *et al.*, 2006. Kinetic evidence for tandemly arranged ligand binding sites in melanocortin 4 receptor complexes, *Neurochem. Int.*

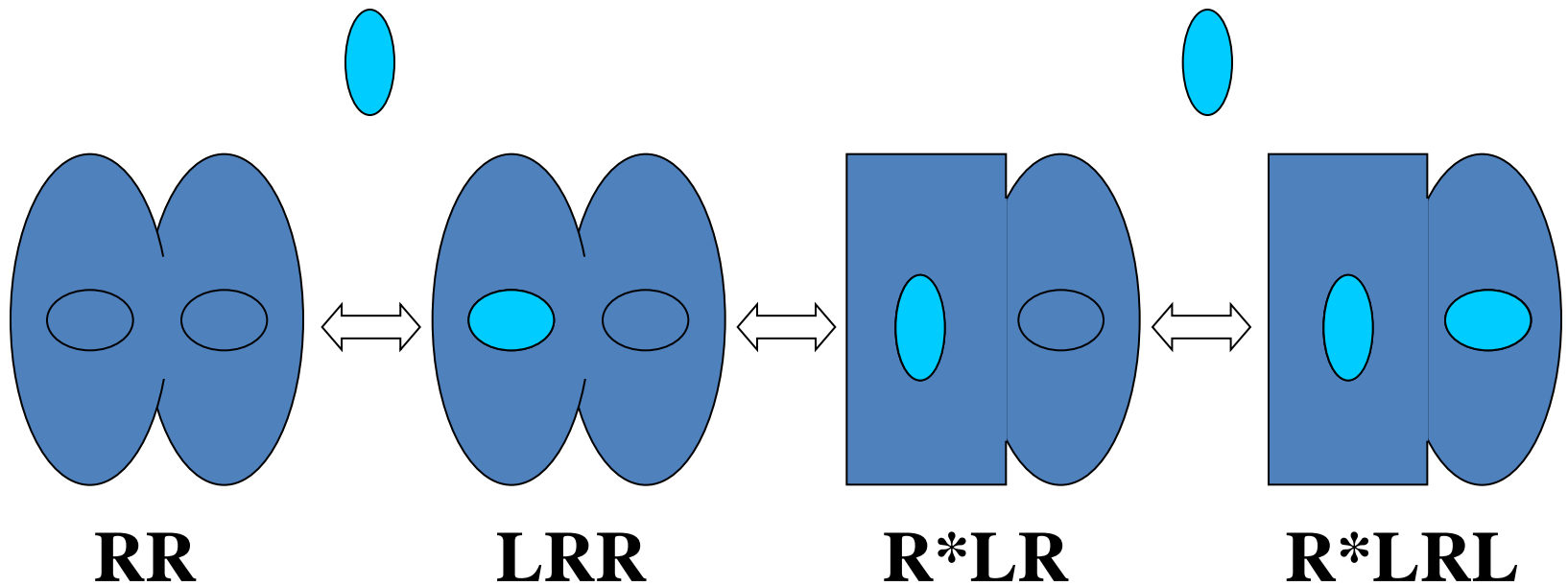
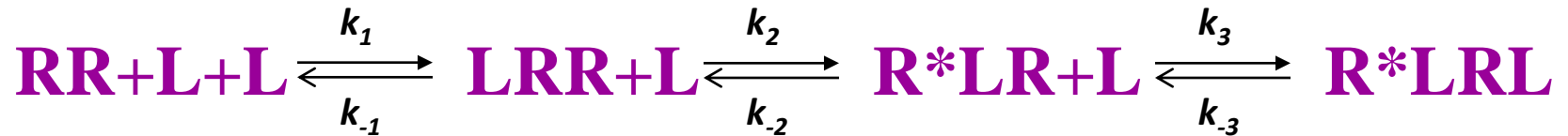
Kopanchuk S., *et al.* 2005, Co-operative regulation of ligand binding to melanocortin receptor subtypes: Evidence for interacting binding sites, *Eur. J. Pharmacol.*

Mutulis F., *et al.*, 2003. A non-peptide radioiodinated high affinity melanocortin-4 receptor ligand. *Journal of Labelled Compounds and Radiopharmaceuticals*

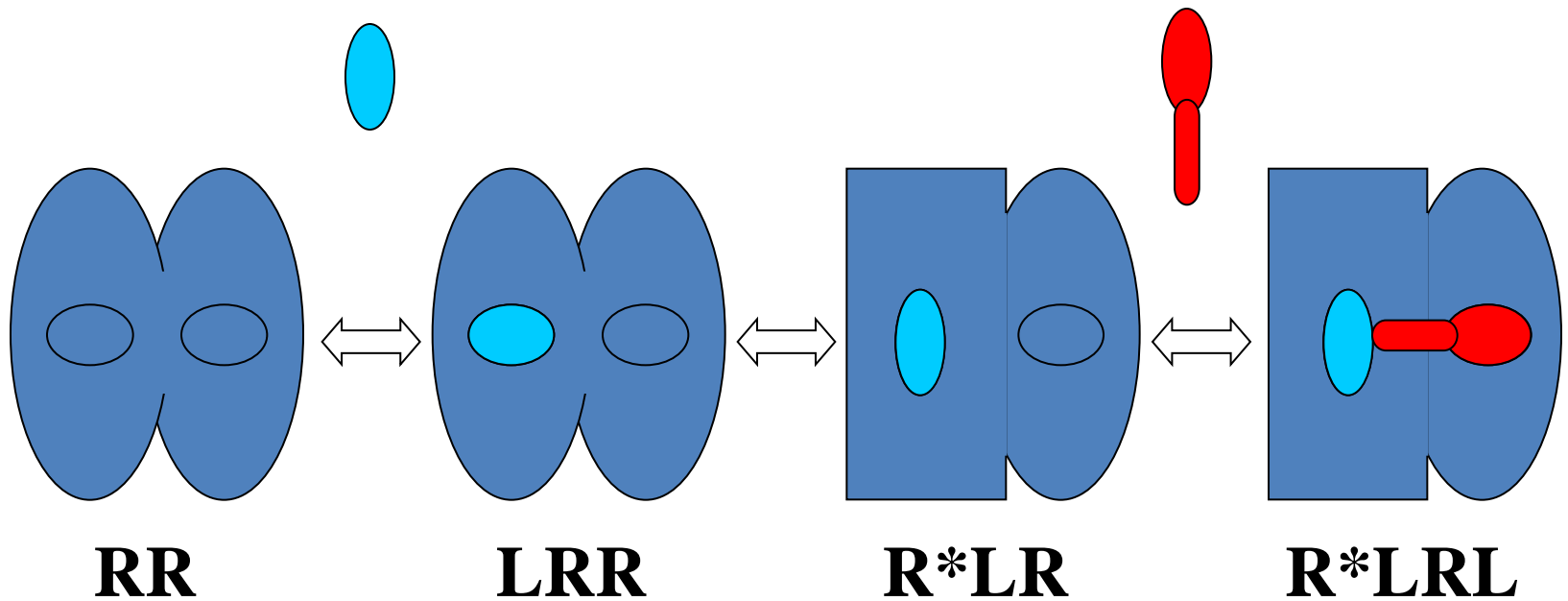
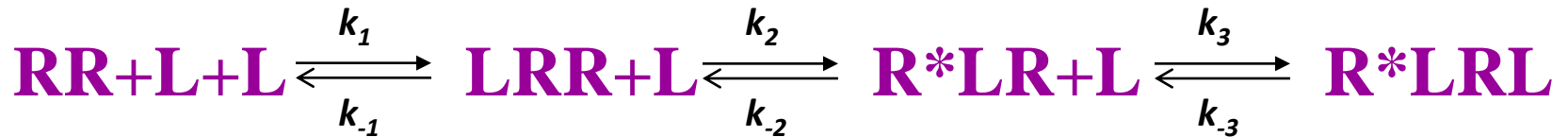
„Two-tandem binding“ model on constitutive dimers



„Two-tandem binding“ model on constitutive dimers



„Two-tandem binding“ model on constitutive dimers

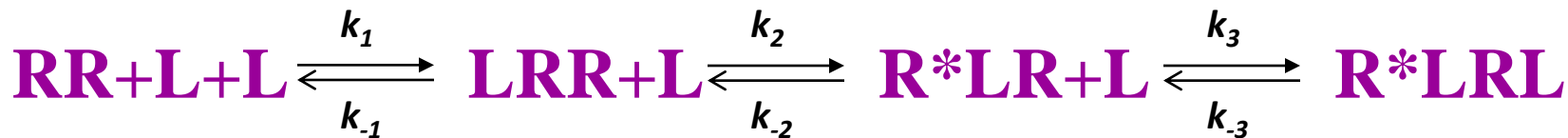
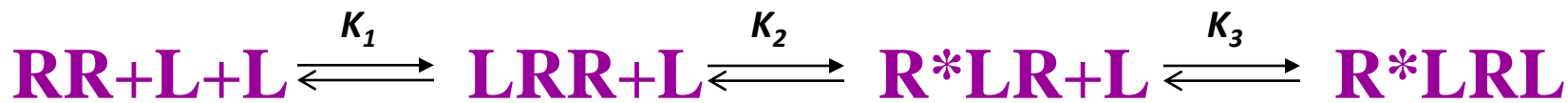


$$rl = \frac{r_o \cdot l \cdot K_2 \cdot K_3}{K_1 \cdot K_2 \cdot K_3 + K_3 \cdot (1 + K_2) \cdot l + l^2} \cdot \left(1 - e^{-\left(\frac{l \cdot (k_{-1} + k_2) \cdot K_2 \cdot K_3}{K_1 \cdot K_2 \cdot K_3 + K_3 \cdot l + l^2} + k_{-1} + k_2 \right) \cdot t} \right)$$

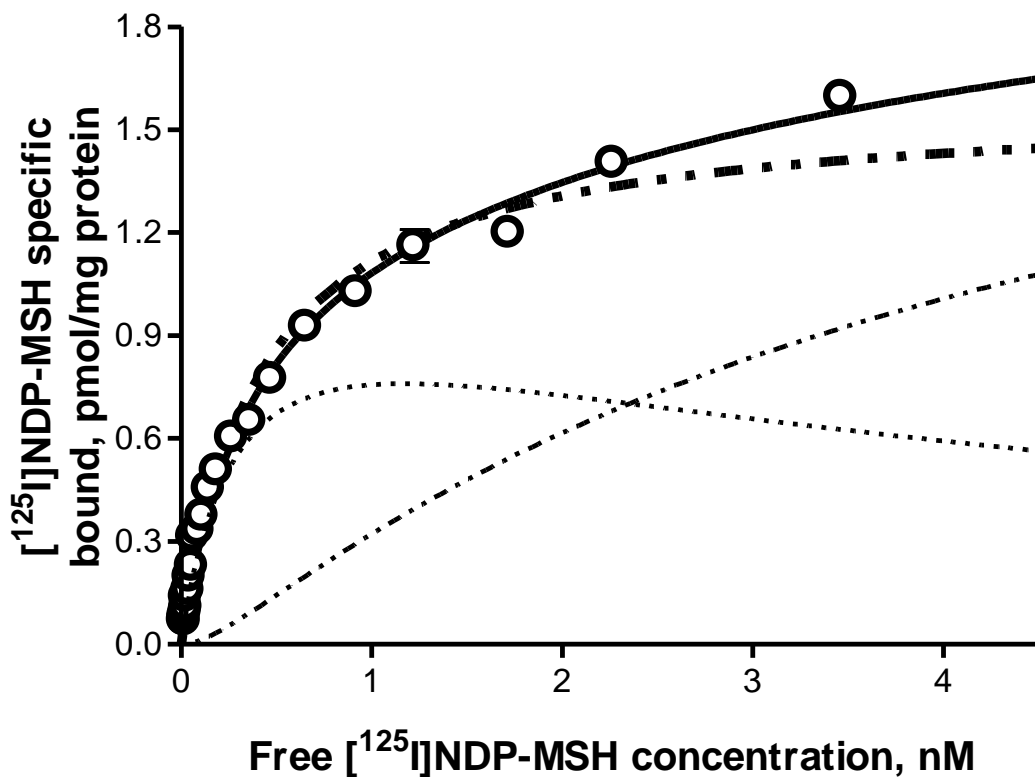
$$lr = \frac{r_o \cdot l \cdot K_3}{K_1 \cdot K_2 \cdot K_3 + K_3 \cdot (1 + K_2) \cdot l + l^2} \cdot \left(1 - e^{-\left(\frac{l \cdot (k_{-2} + k_3 \cdot l) \cdot K_3}{K_1 \cdot K_2 \cdot K_3 + K_2 \cdot K_3 \cdot l + l^2} + k_3 \cdot l + k_{-2} \right) \cdot t} \right)$$

$$lrl = \frac{r_o \cdot l^2}{K_1 \cdot K_2 \cdot K_3 + K_3 \cdot (1 + K_2) \cdot l + l^2} \cdot \left(1 - e^{-\left(\frac{k_3 \cdot l^2}{K_1 \cdot K_2 + l \cdot (1 + K_2)} + k_{-3} \right) \cdot t} \right)$$

$$\text{Bound} = rl + lr + 2 \cdot lrl$$



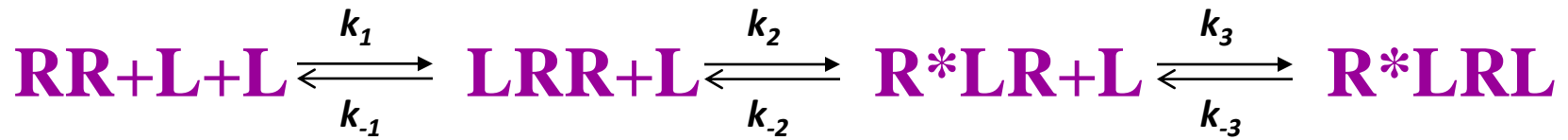
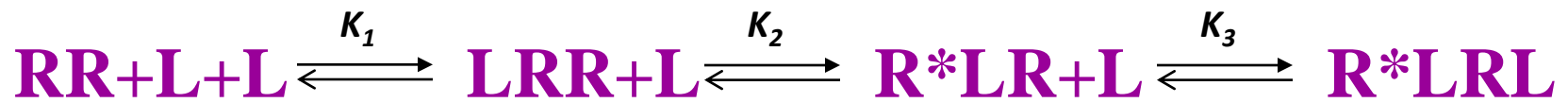
$t \rightarrow \infty$



$$B = \frac{r_o \cdot l}{\frac{K_{all} - l^2}{K_{add} + 2 \cdot l} + l}$$

$$K_{all} = K_1 \cdot K_2 \cdot K_3$$

$$K_{add} = K_3 (1 + K_2)$$



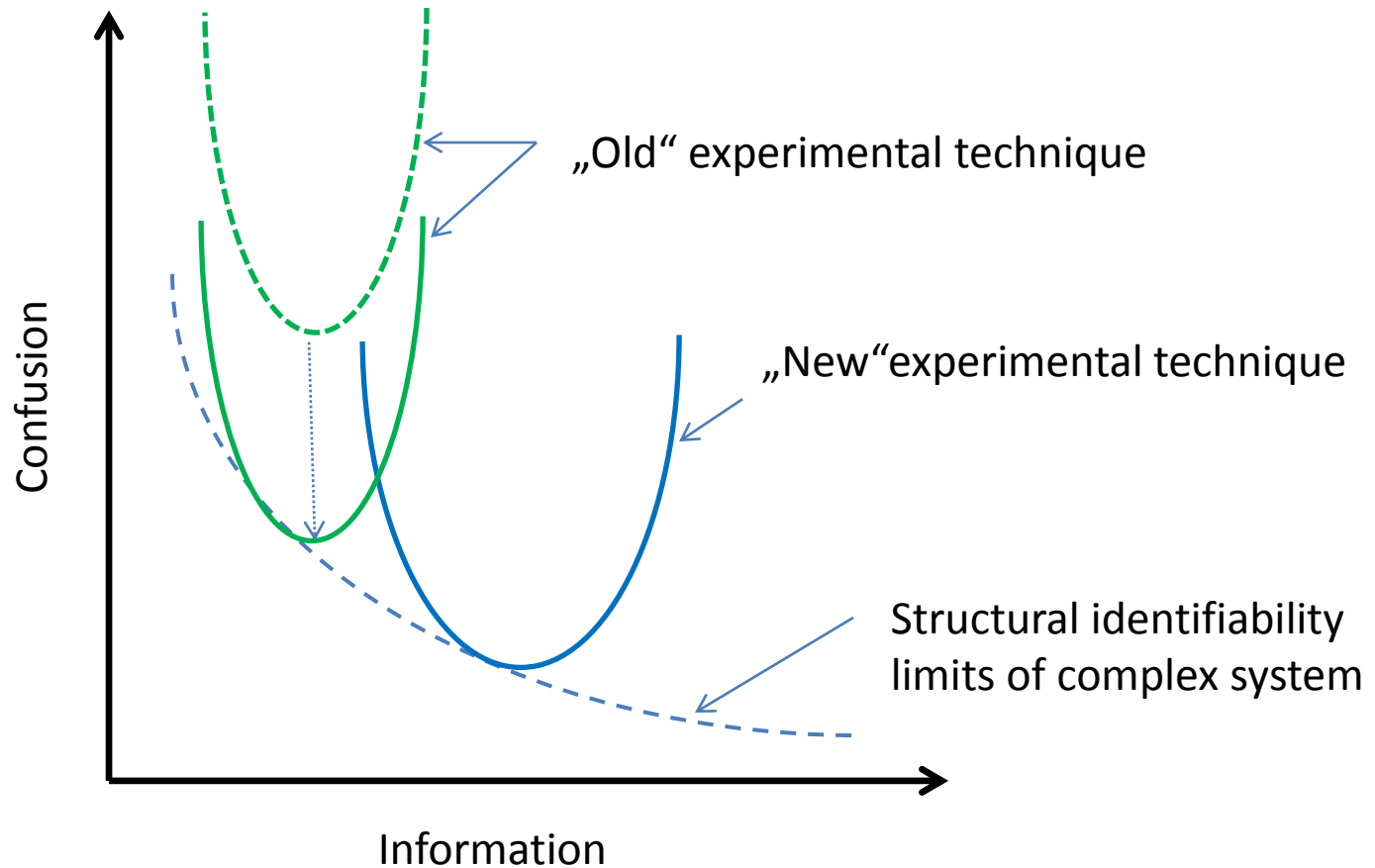
$$B = \frac{r_o \cdot l}{K(L) + l} \quad \longleftarrow \quad B = \frac{r_o \cdot l}{\frac{K_{all} - l^2}{K_{add} + 2 \cdot l} + l}$$

$$\frac{dK(L)}{dL} = \kappa$$

$$K_{all} = K_1 \cdot K_2 \cdot K_3$$

$$K_{add} = K_3 (1 + K_2)$$

Our confusion as function of available information with practical and structural identifiability limits



“To link or not to link: the “global” question”

Total Fluorescence Intensity (TFI) $\sim I_x + I_y + I_z$

"sometimes" (isotropic environment): $I_x = I_y = I_{\text{per}}$ and $\neq I_z = I_{\text{paral}}$

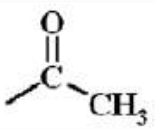
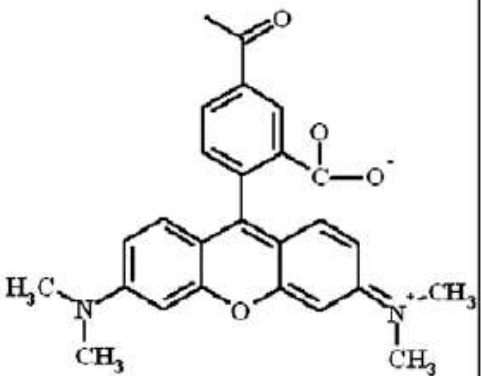
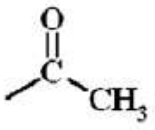
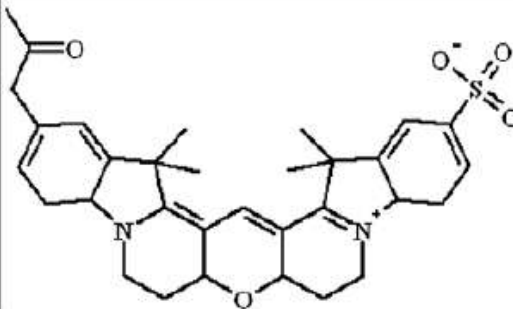
Detected Fluorescence Intensity (DFI) $\sim I_{\text{paral}} + I_{\text{per}}$ but $TFI \sim I_{\text{per}} + 2 \cdot I_{\text{paral}}$

Fluorescence Anisotropy (FA)

$$r = \frac{I_{\text{paral}} - I_{\text{per}}}{I_{\text{paral}} + 2 \cdot I_{\text{per}}}$$

$$TFI = \frac{3 \cdot DFI}{2 + r}$$



	R_1	R_2
NDP- α -MSH		- H
TAMRA-NDP- α -MSH		- H
Cy3B-NDP- α -MSH		

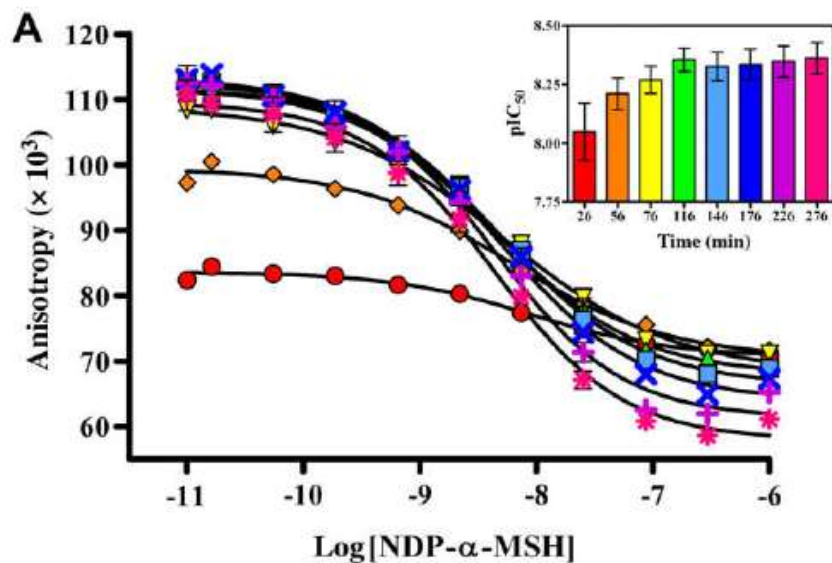
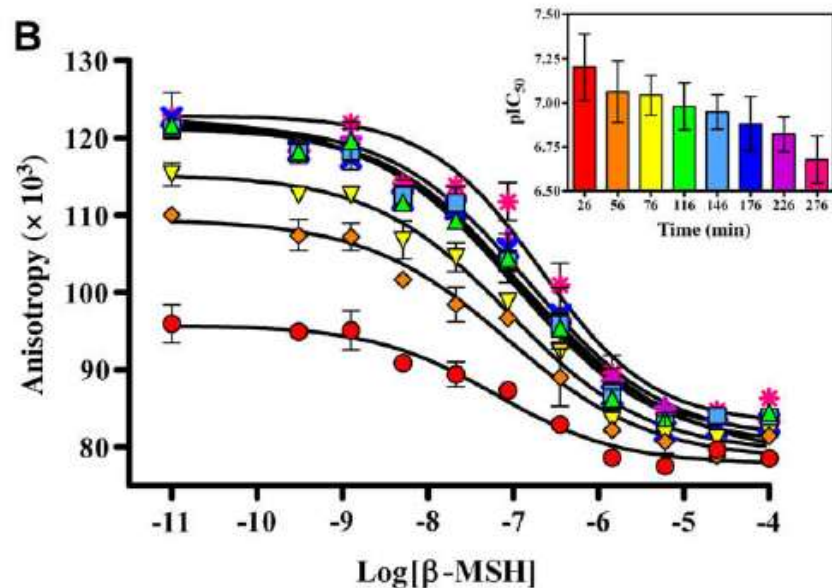


Table 1

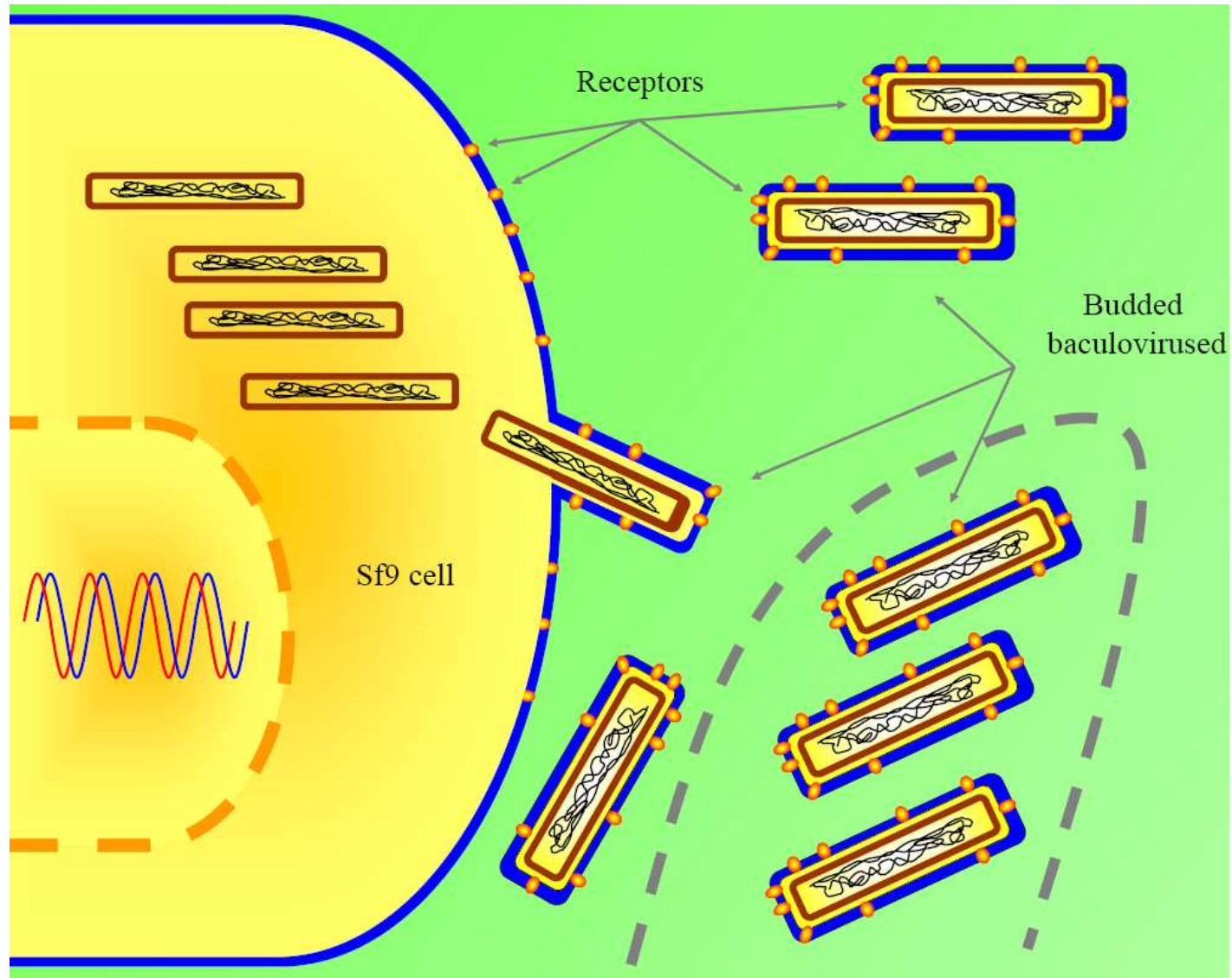
Apparent potencies of ligands to MC₄ receptors in Sf9 cell membranes determined in competition with Cy3B-NDP- α -MSH and TAMRA-NDP- α -MSH.

	Cy3B-NDP- α -MSH		TAMRA-NDP- α -MSH	
	pIC ₅₀ ^a	Hill slope	pIC ₅₀ ^a	Hill slope
NDP- α -MSH	8.35 \pm 0.12	0.80 \pm 0.18	8.14 \pm 0.09	0.92 \pm 0.15
β -MSH	6.55 \pm 0.58	0.53 \pm 0.08	6.14 \pm 0.10	1.08 \pm 0.21
HS-024	7.79 \pm 0.12	0.97 \pm 0.31	7.62 \pm 0.10	1.08 \pm 0.24
I-THIQ	7.59 \pm 0.12	0.73 \pm 0.15	7.69 \pm 0.03	1.04 \pm 0.08

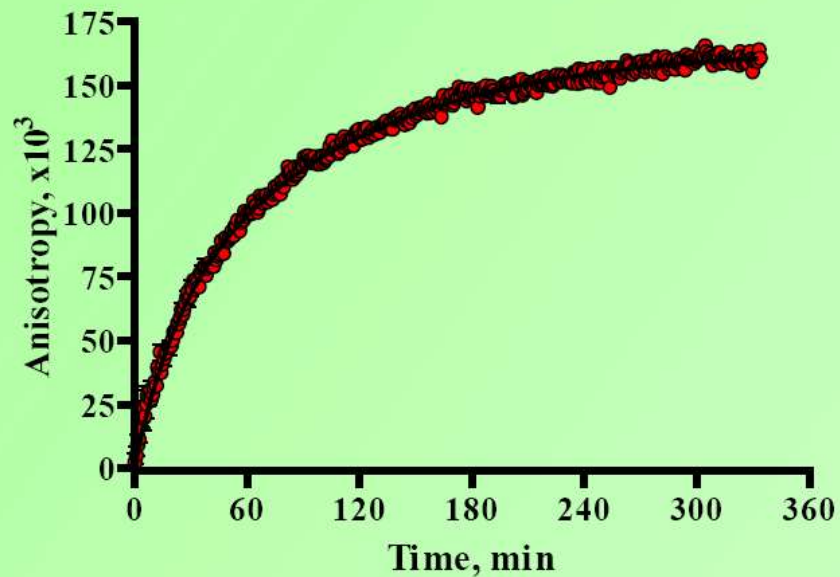


Veiksina S., et al. 2010. Fluorescence anisotropy assays for pharmacological characterisation of ligand binding dynamics to melanocortin 4 receptors. *Analytical Biochemistry*

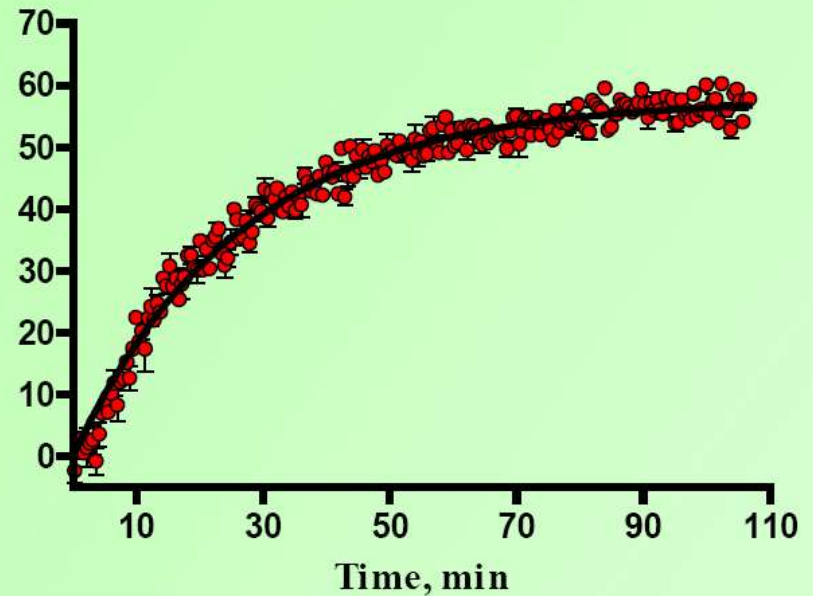
Baculovirus Surface Display System



Time course of specific changes in fluorescence anisotropy caused
by 1 nM Cy3B-NDP- α -MSH binding to MC₄ receptors

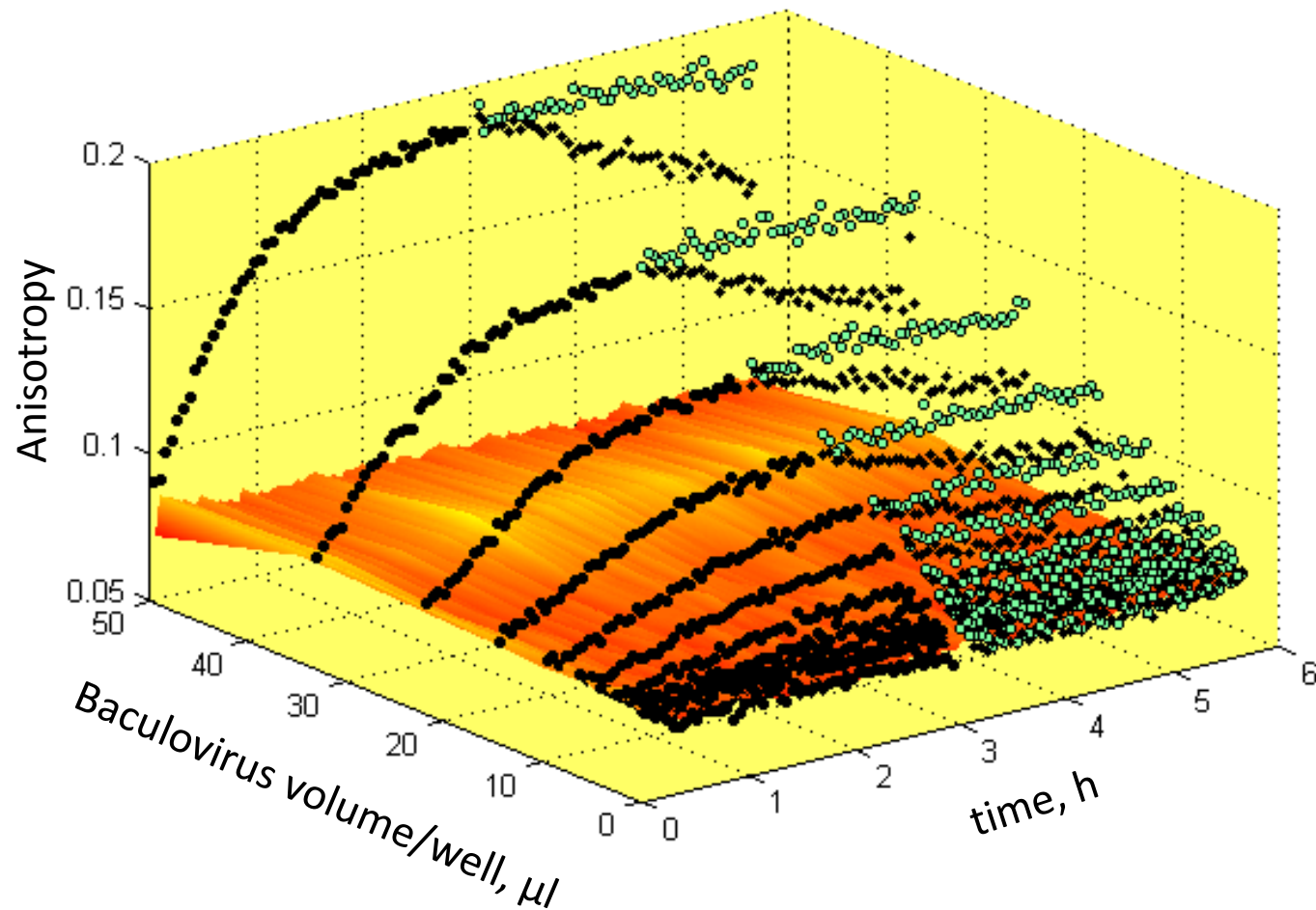


Baculovirus assay

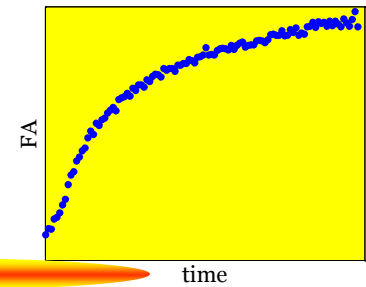
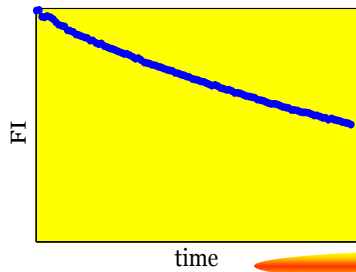


Membrane homogenate

Time course of changes in fluorescence anisotropy caused by 1 nM
Cy3B-NDP- α -MSH binding to MC₄ receptors



Multivariate Global Data Analysis (based on the mechanistic models)



Initial state values. Events. Parameter values

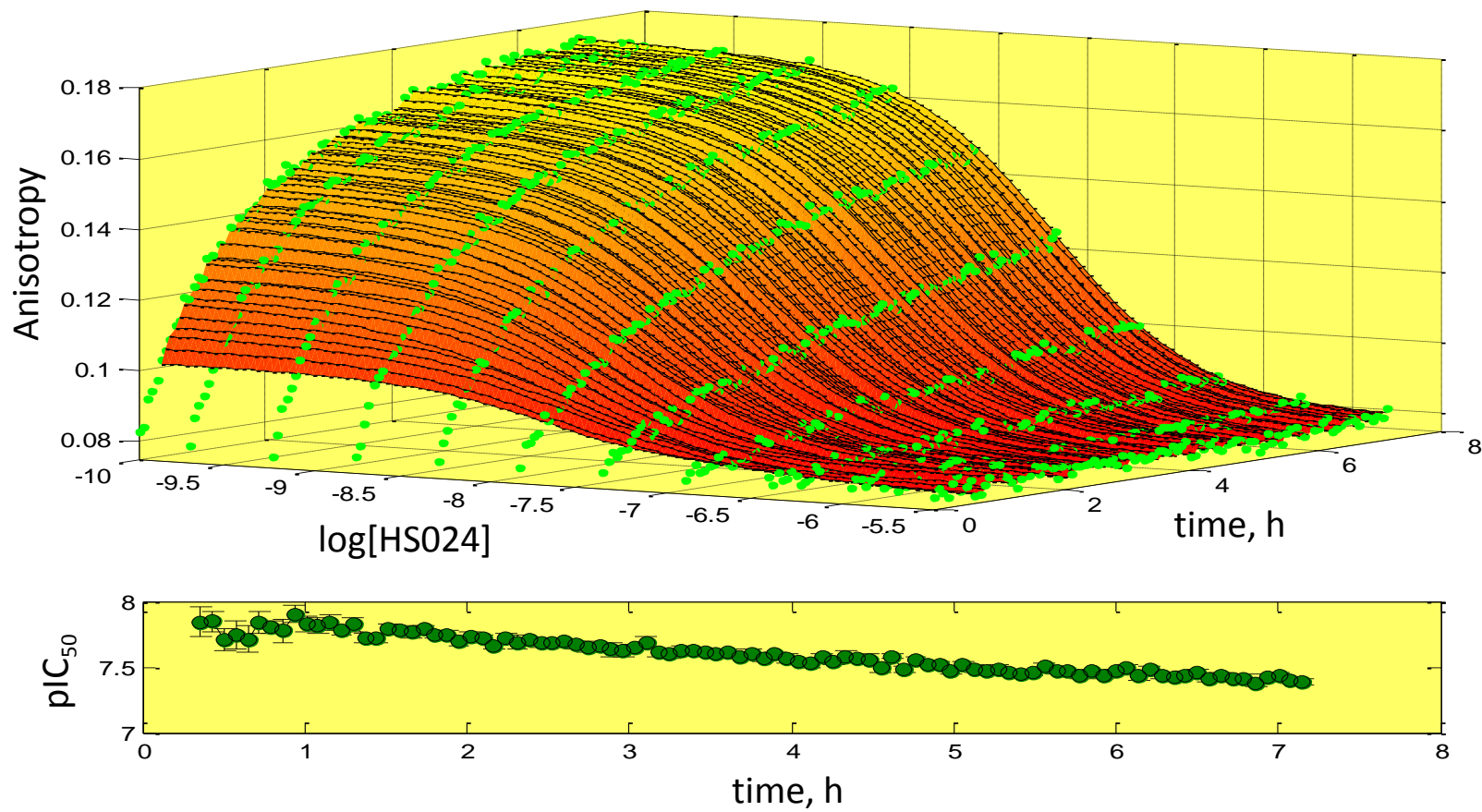
Fractions of each state are generated by numerical integrations with ODE solvers

Calculated $FI(t)$

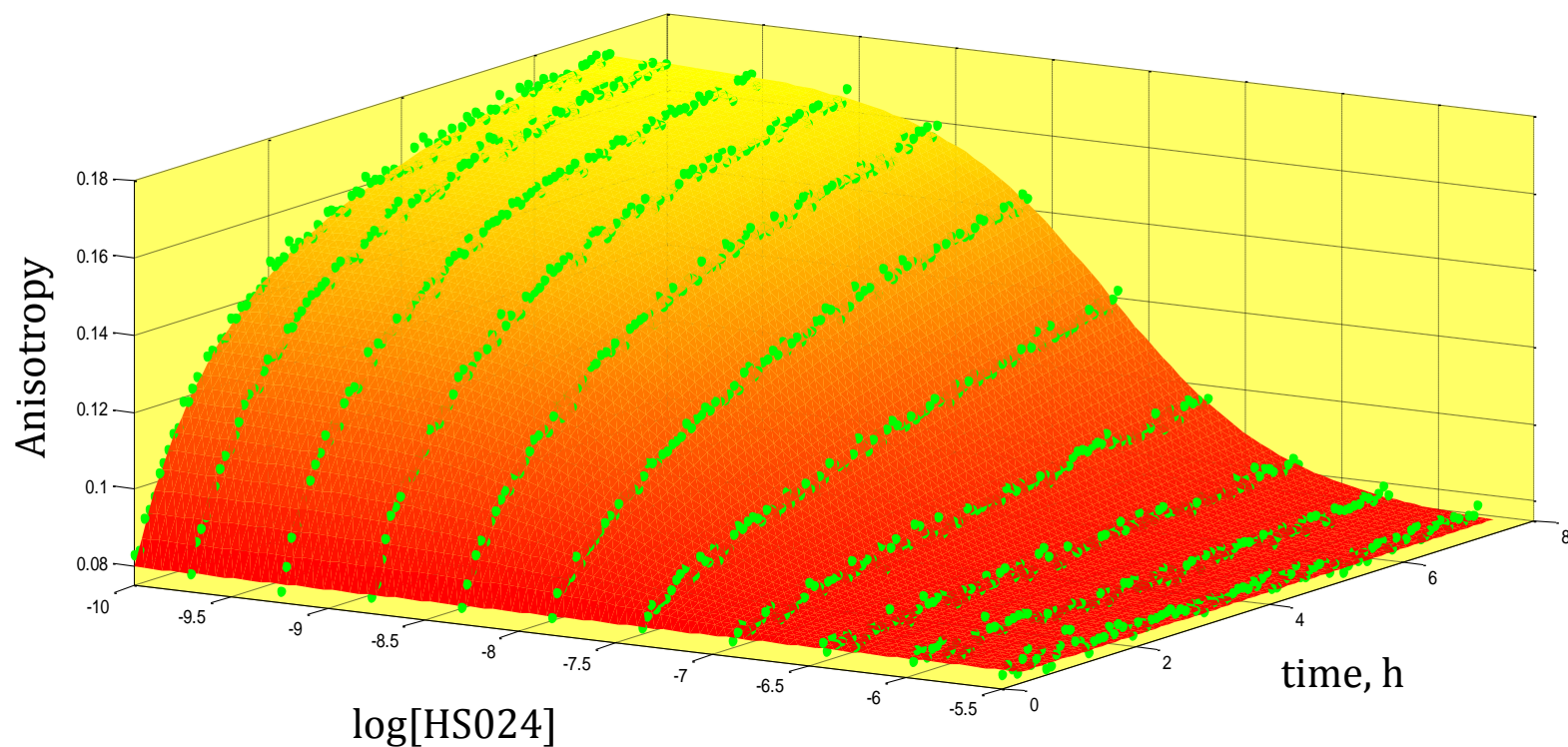
Calculated $FA(t)$

Parameter estimation with global stochastic optimisation algorithms (Differential Evolution, Adaptive Particle Swarm, etc) and fitting (Nelder-Mead nonlinear simplex) for internally consistent solution

Time courses of changes in fluorescence anisotropy caused by Cy3B labeled NDP- α -MSH binding to MC₄ receptors exposed on baculovirus surface in the presence of different concentrations of HS024

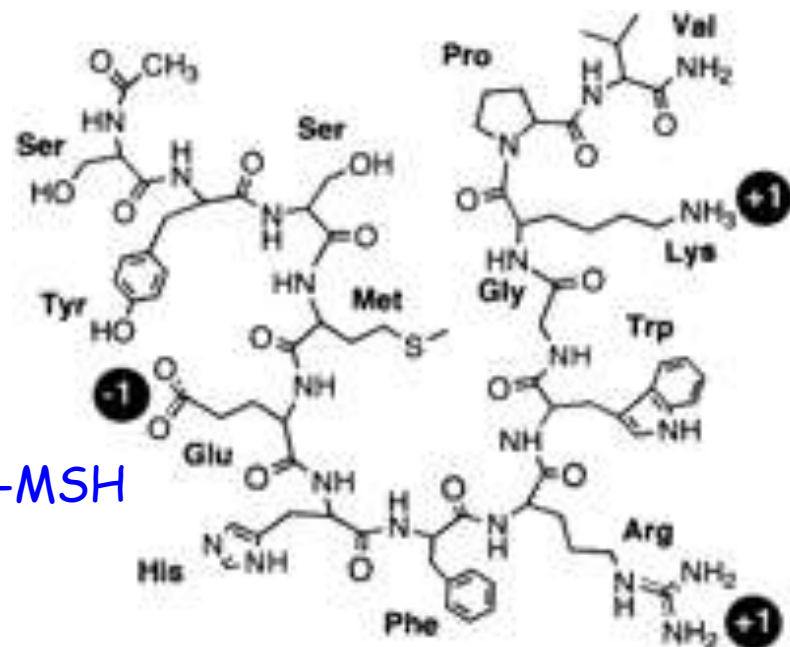


Time courses of changes in fluorescence anisotropy caused by Cy3B labeled NDP- α -MSH binding to MC₄ receptors exposed on baculovirus surface in the presence of different concentrations of HS024

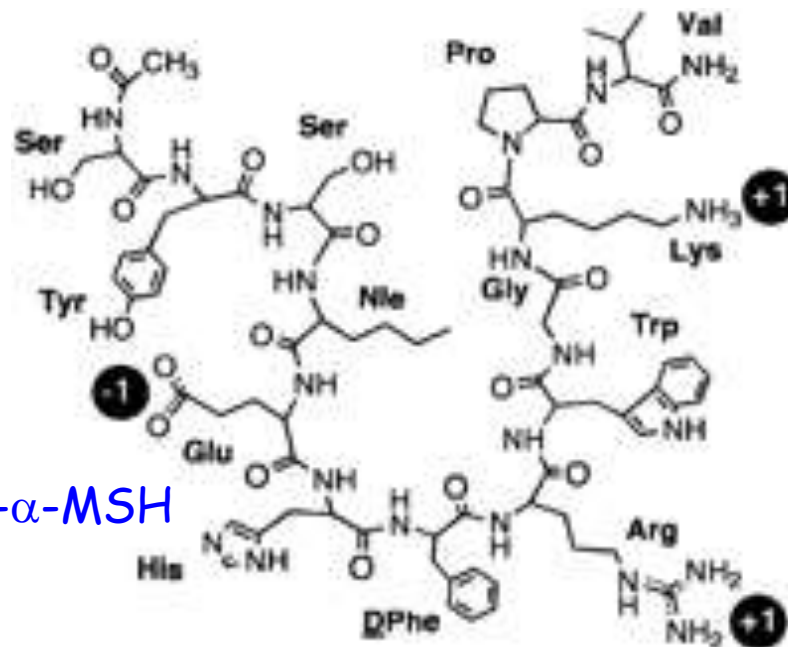


Melanocortin peptides

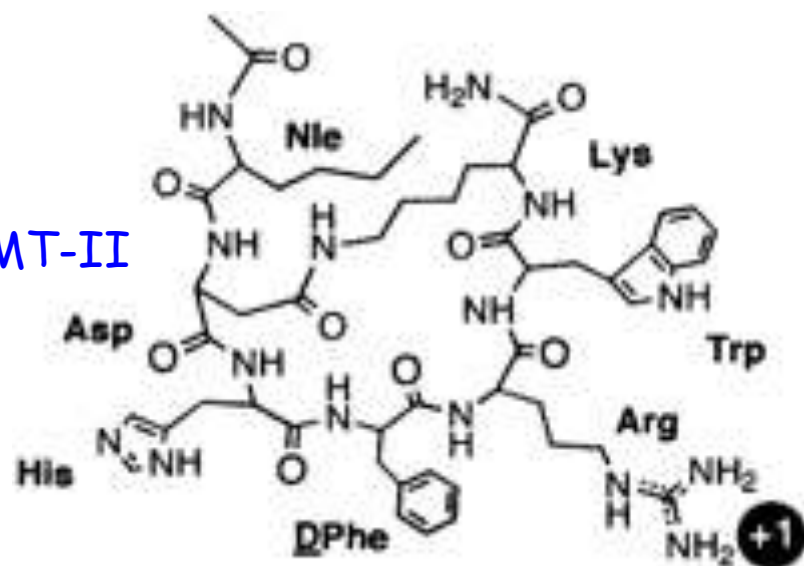
α -MSH



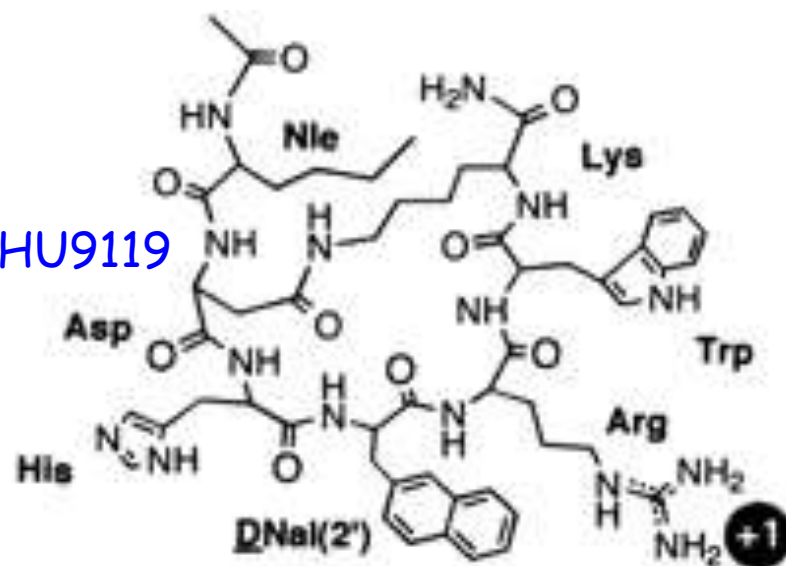
NDP- α -MSH



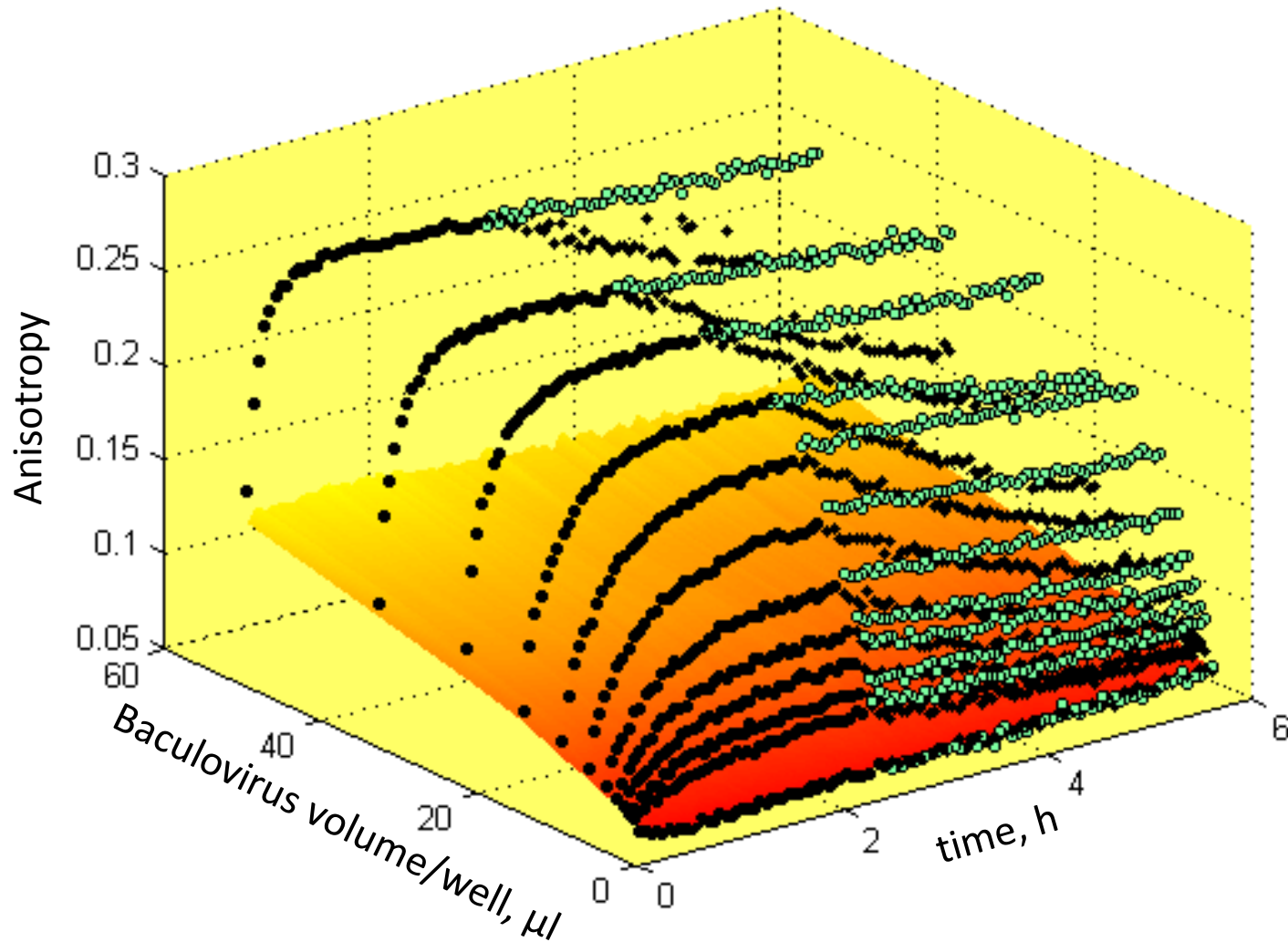
MT-II



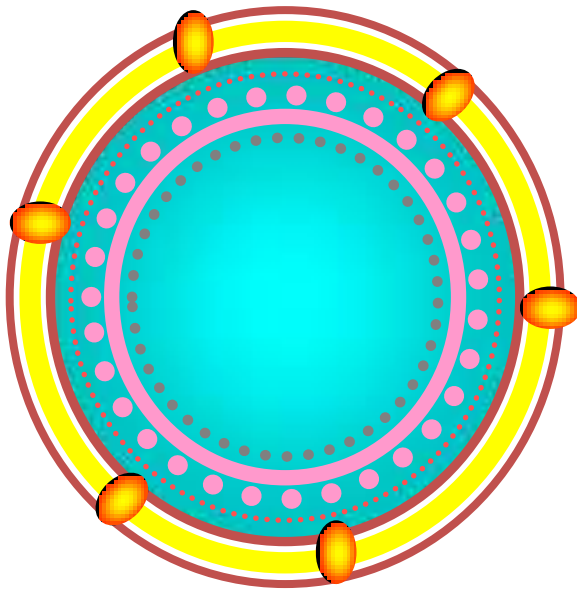
SHU9119



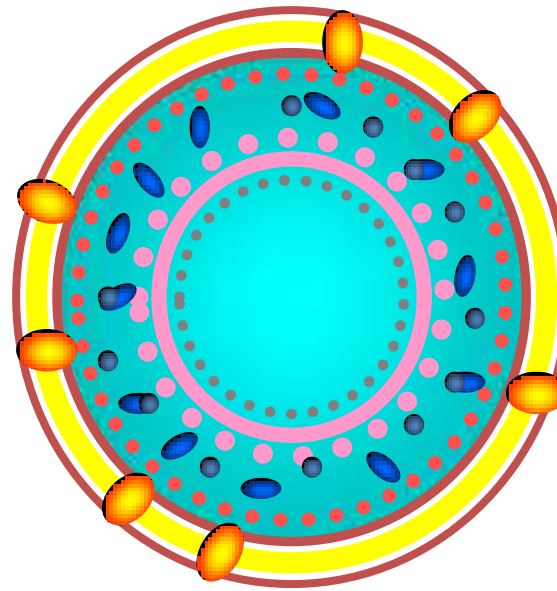
Time course of changes in fluorescence anisotropy caused by 1 nM Cy3B-Peptide_1 (A modification) binding to MC₄ receptors



Receptor displayed on the surface of retrovirus like particle (VLP) (*mammalian system*)



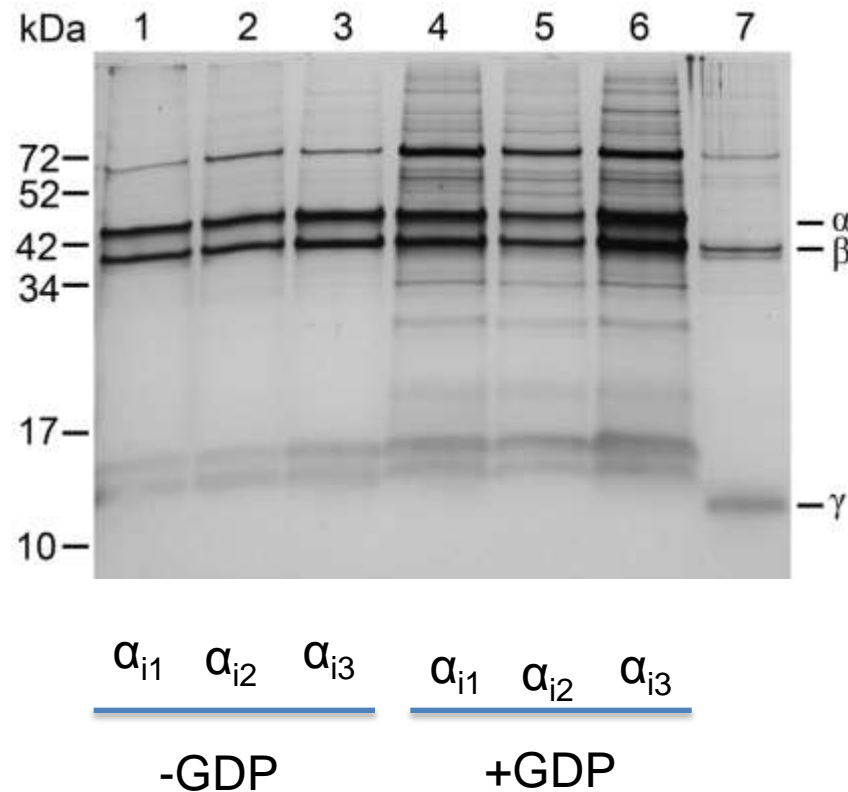
Receptors on VLPs



Receptors and other molecules of signaling mashinery

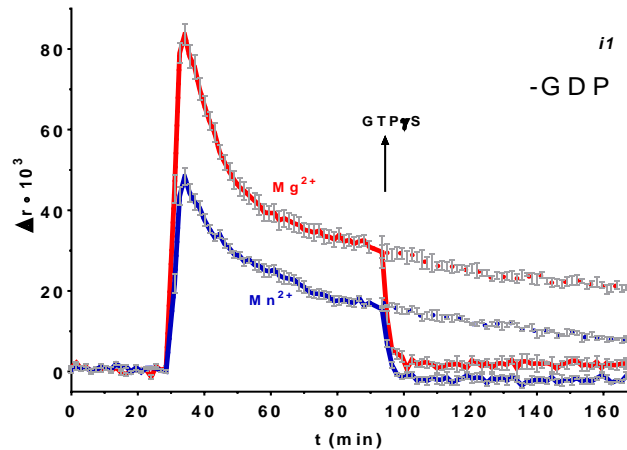


Recombinant heterotrimeric G-protein α_{i1} , α_{i2} and α_{i3} subunits were purified in GDP-depleting conditions by affinity chromatography using StreptII-tagged $\beta_1\gamma_2$ subunits

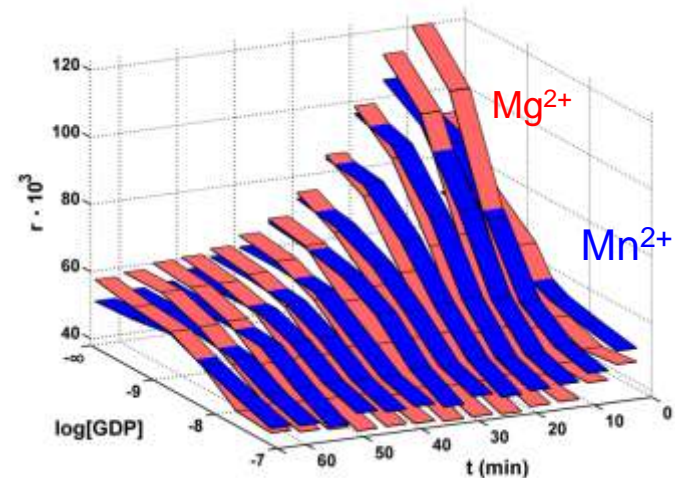
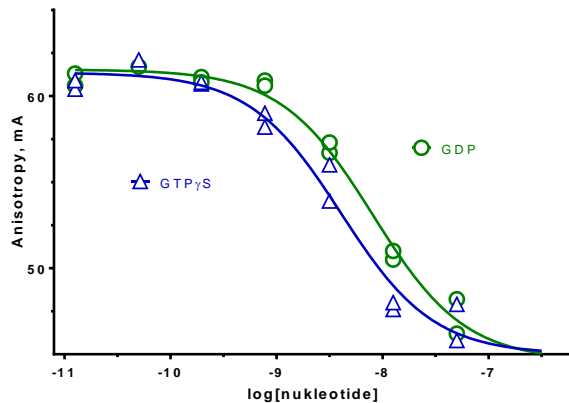
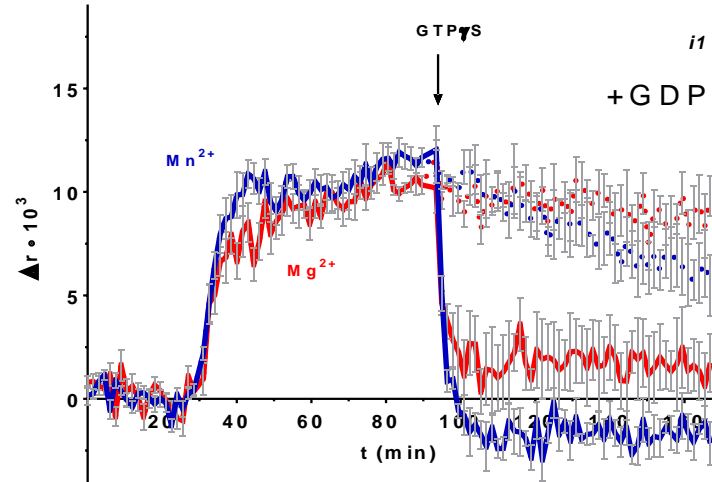


Tõnson L., et al. 2012. Characterization of heterotrimeric nucleotide-depleted G α_i -proteins by Bodipy-FL-GTP γ S fluorescence anisotropy. *Archives of Biochemistry and Biophysics*

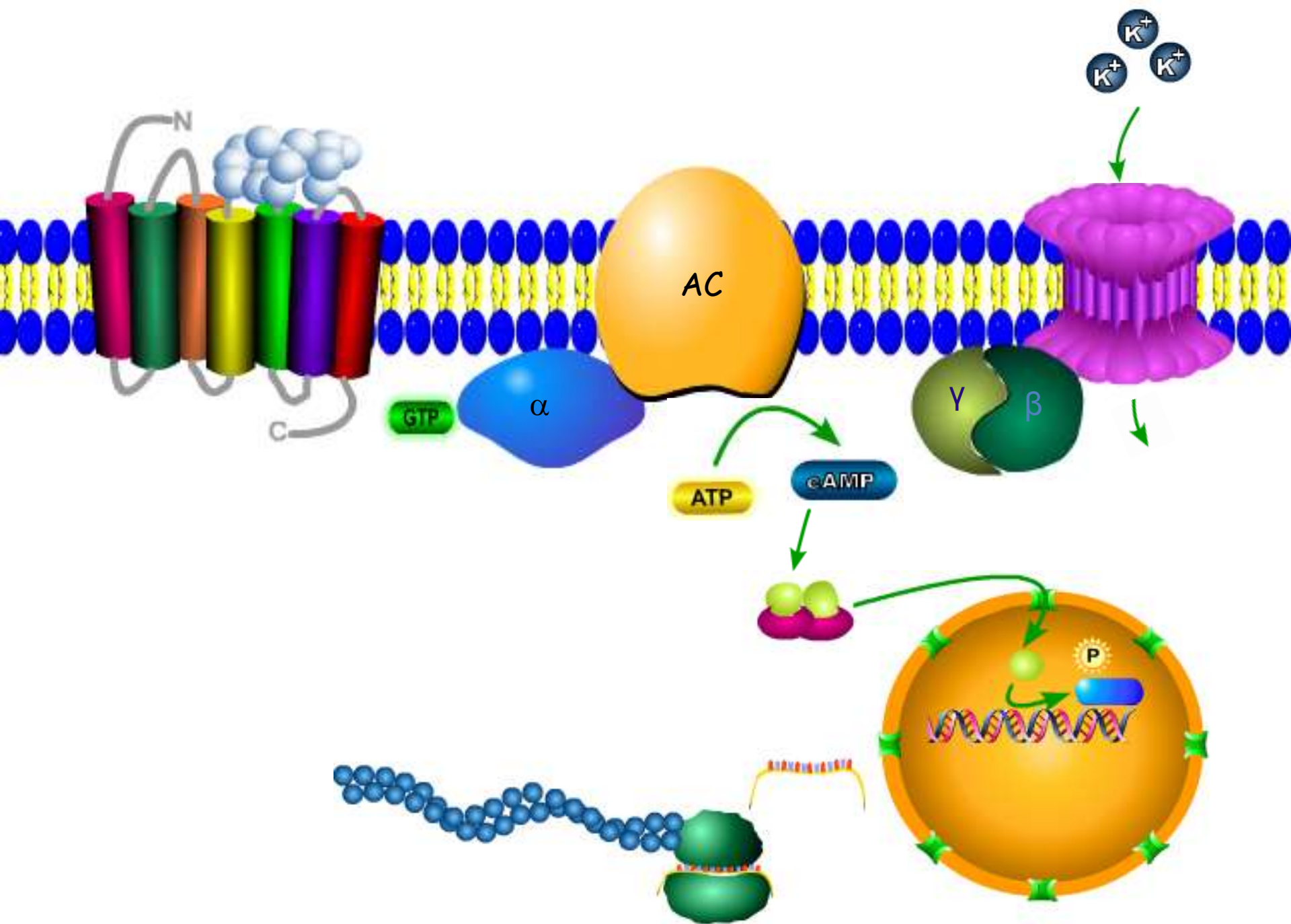
Association and dissociation kinetics of Bodipy-FL-GTP γ S binding to G-protein heterotrimers



GDP depleted G protein

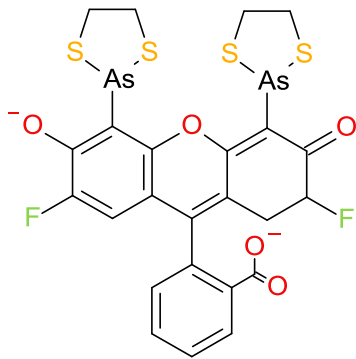


Tõnson L., et al. 2012. Characterization of heterotrimeric nucleotide-depleted G α i-proteins by Bodipy-FL-GTP γ S fluorescence anisotropy. *Archives of Biochemistry and Biophysics*

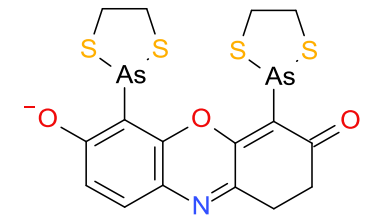
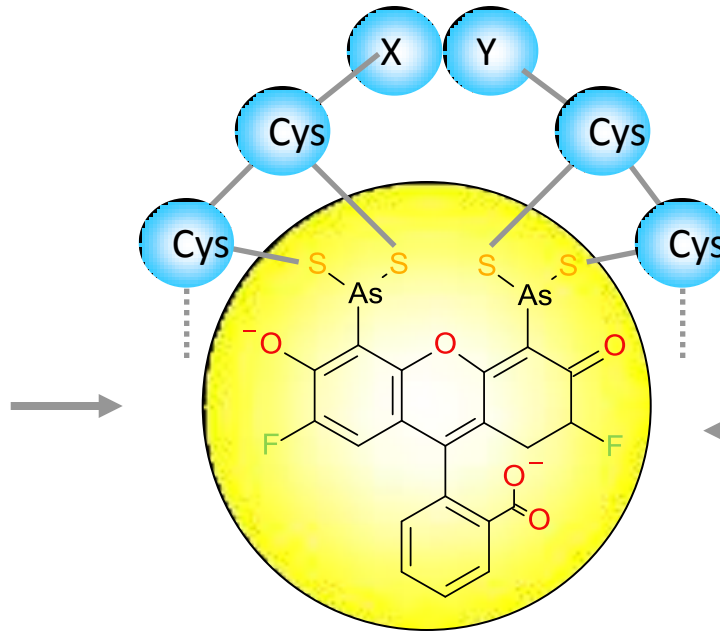


Fluorescently labeled G-proteins: α_s , α_{i2} , α_q , γ_1

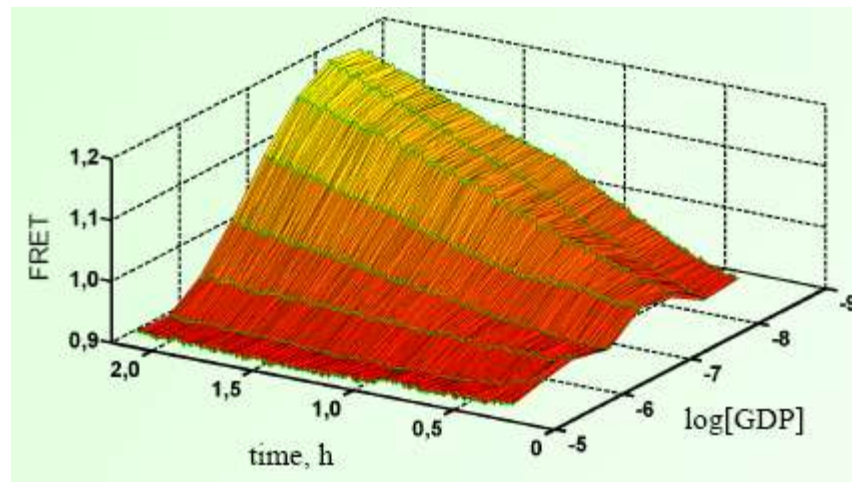
X = Pro or Lys
Y = Gly or Ala



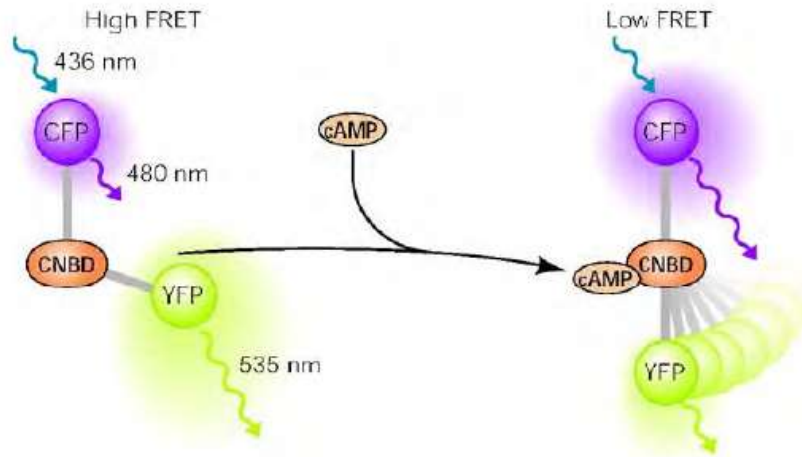
F2FlAsH



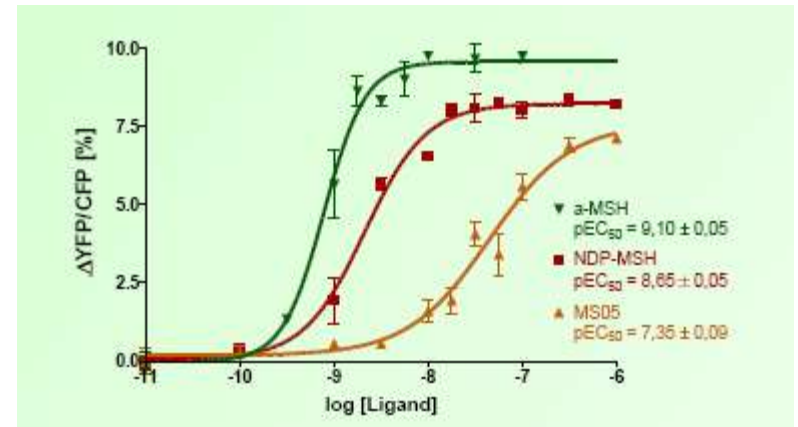
ReAsH



BacMam expression system is suitable for expression of cAMP-sensor protein (Epac-camps) in mammalian cells

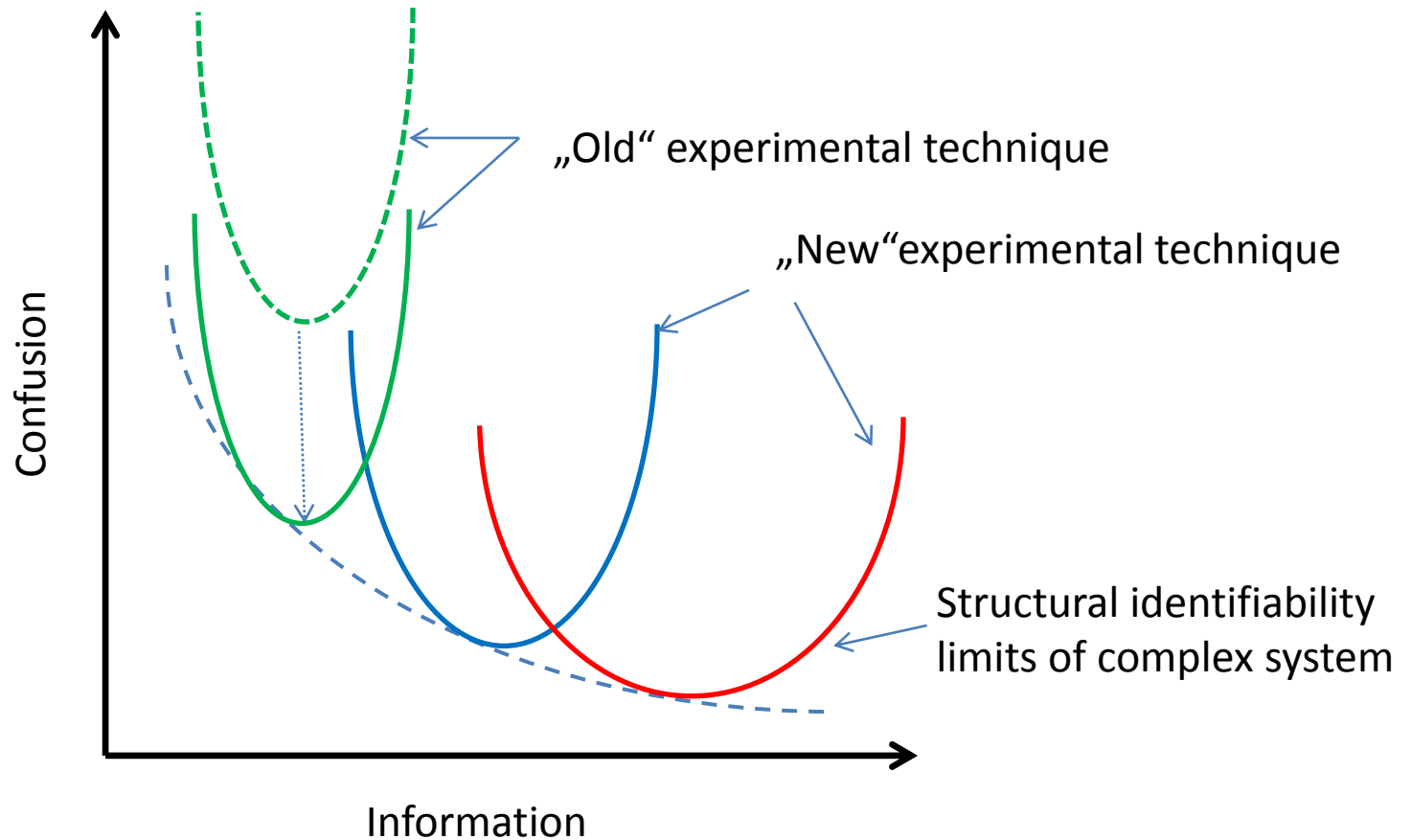


Nikolaev et al. (2004)

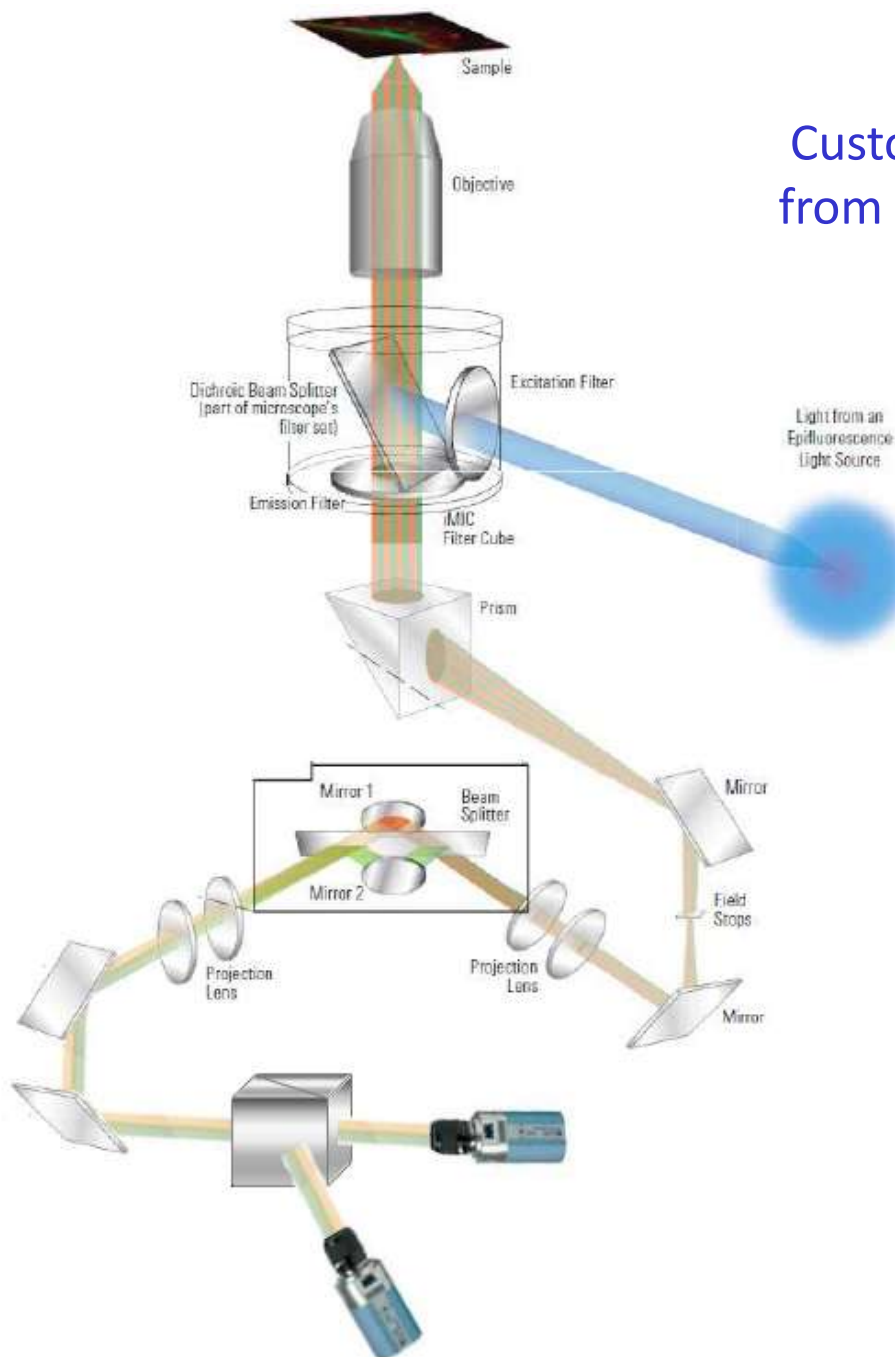


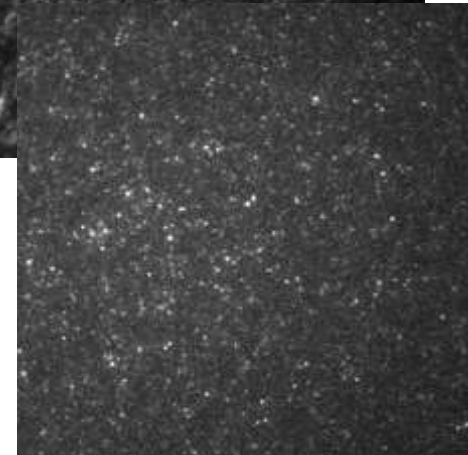
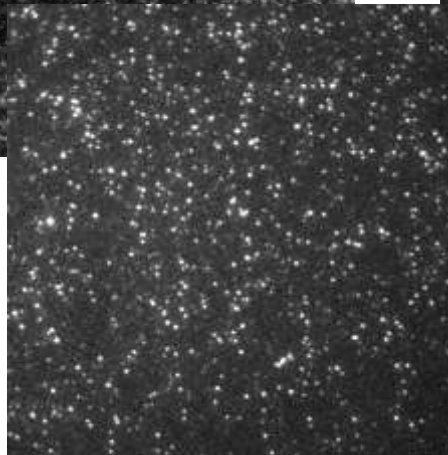
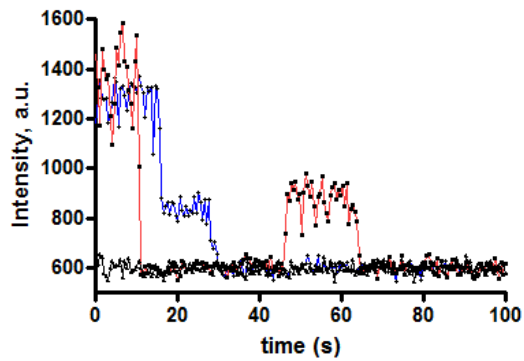
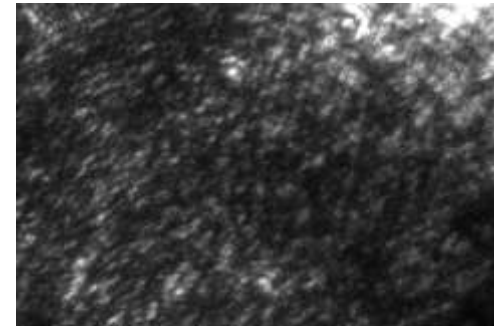
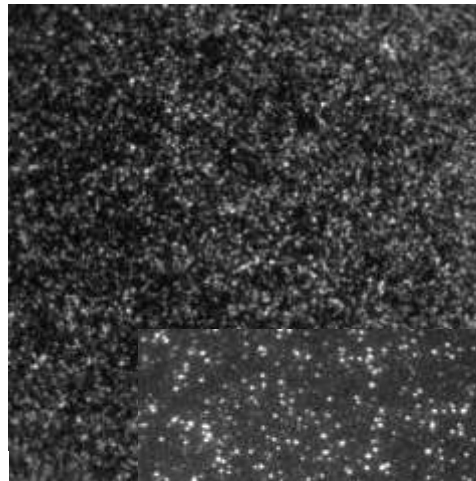
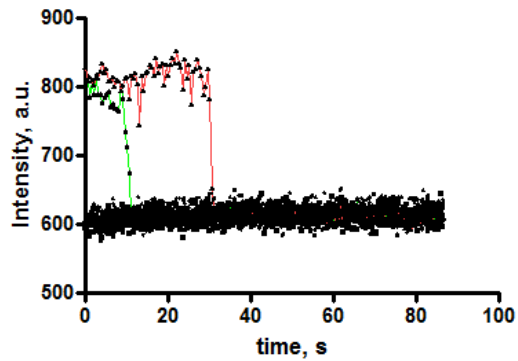
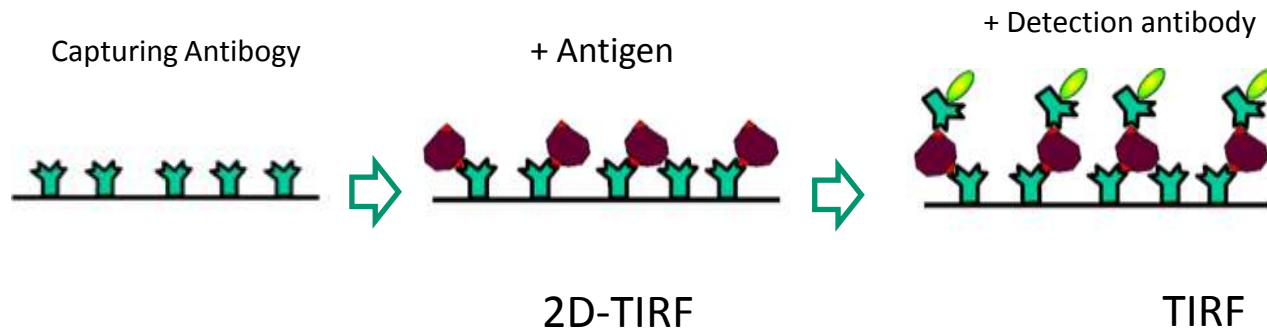
Mazina, O., et al. 2012. BacMam system for FRET based cAMP sensor expression in studies of melanocortin MC1 receptor activation. *Journal of Biomolecular Screening*

Our confusion as function of available information with practical and structural identifiability limits



Custom made iMIC TIRF digital microscope
from Till Photonics with Azzam polarimeter
architecture in the emission path





1 pM Atto532-X

Control

Thank you for your attention!

